

## Reference Protocol for Anti-Lamin B1 (HS-404 013) Western Blots with Alkaline Phosphatase Detection

## Materials and Reagents

- Ponceau S staining solution: 5% acetic acid, 0.1% Ponceau S
- 5% skimmed milk-TBST: 20 mM Tris-HCl, pH 7.5; 150 mM NaCl; 5% (w/v) skimmed milk powder; 0.02% sodium azide; 0.1% Tween 20
- Secondary antibody: AP-conjugated anti-rabbit secondary antibody
- Substrat buffer for alkaline phosphatase: 100 mM Tris-HCl, pH 9.5; 100 mM NaCl; 5 mM MgCl<sub>2</sub>
- BCIP staining solution: 20 mg/ml in 100% di-methyl formamide
- NBT staining solution: 50 mg/ml in 70% di-methyl formamide
- **Staining solution complete:** Substrate buffer containing 80 μl BCIP solution and 60 μl NBT solution per 10 ml. Always prepare this solution **fresh** shortly before use.

## **Procedure**

The protein sample to be examined and a molecular weight standard are separated by SDS PAGE and transferred to a nitrocellulose membrane by electro-blotting. Follow the manufacturer's instructions for your SDS-PAGE and blotting device.

- 1. Stain the membrane with **Ponceau S staining solution** for several minutes at room temperature to check transfer.
- 2. Rinse the membrane in water to remove the Ponceau staining and incubate in **5% skimmed milk-TBST** for 30 min on a lab shaker.
- 3. Replace with fresh 5% skimmed milk-TBST containing the primary antibody at a 1:1000 dilution and incubate for at least 2 h at RT on a lab shaker or over-night at 4°C.
- 4. Wash 3 4 times with 5% skimmed milk-TBST for 10 min each time.
- 5. Replace with fresh 5% skimmed milk-TBST containing AP-conjugated secondary antibody diluted to the manufacturer's recommended concentration and incubate for at least 1 h on a lab shaker.
- 6. Wash 3 times with 5% skimmed milk-TBST for 10 min each time.
- 7. Wash with **substrate buffer** and let equilibrate for 5 min.



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- 8. Replace with fresh **staining solution complete** and develop for 15 30 min. Time can be shortened or extended, if signals are extremely strong or weak, respectively.
- 9. Stop staining reaction by washing 3 times with H<sub>2</sub>O.

**Note:** The SYSY standard protocol generates good staining results in the SYSY labs and may be used as a reference. However, to achieve the highest specific signal and lowest non-specific background signal, the best antibody concentration, incubation temperature and incubation time for each antibody must be individually determined. Please also refer to our general protocols.