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Reference Protocol for Anti-IBA1 (HS-234 308) Immunohistochemistry using DAB as Chromogen

Tissue Fixation

+ 3.7% formaldehyde (24 h), 3.5 μM paraffin sections

Materials and Reagents

| Food Steamer | Braun, Multigourmet | | |
|--|---------------------|--------------|--|
| Staining Containers with slide holders (e.g. Tissue-Tek) | | | |
| Protein Block, Serum-Free | Agilent | X0909 | |
| Antibody diluent | Agilent | S2022 | |
| Biotinylated anti-Guinea pig antibody | Jackson | 106-065-003 | |
| ABC HRP Kit, Standard | Vectorlabs | PK-4000 | |
| ImmPACT DAB | Vectorlabs | SK-4105 | |
| Hydrogen peroxide 30% | Merck | 1.07298.0250 | |
| • PBS (pH 7.4) | | | |
| • TBST (TBS, 0.05% Tween 20, pH 7.6) | | | |
| Antigen Retrieval buffer: Citrate Buffer (10 mM Citrate, 0.05% Tween 20, pH 6.0) | | | |
| • Xylene, 100% ethanol, 90% ethanol, 80% ethanol and 70% ethanol, 2-propanol | | | |
| Optional: Hematoxylin Solution (Mayer's, Modified) or other nuclear counterstain | | | |
| Optional: Avidin/Biotin Blocking Kit | Vectorlabs SP-2001 | | |

• Non-aqueous mounting medium

Method

1. Deparaffinize and hydrate tissue sections

| a. | Xylol | 2 x 5 min |
|----|-----------------|------------|
| b. | 100% EtOH | 2 x 2 min |
| c. | 90% EtOH | 1 x 2 min |
| d. | 80% EtOH | 1 x 2 min |
| e. | 70% EtOH | 2 x 2 min |
| f. | Deionized Water | 1 x 20 sec |
| | | |

g. PBS 1 x 2 min

Keep the slides in PBS until ready to perform the Antigen Retrieval. Do not allow the slides to dry out

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2. Antigen Retrieval (AR) using a food steamer

- a. Heat the steamer with a suitable staining container filled with Antigen Retrieval buffer to ~97°C
- b. Transfer the sections into the staining box, wait until the temperature reaches 97°C
- c. Incubate the sections in the steamer for 30 min
- Remove the staining container from the steamer and allow the slides to cool down for 20 min (target end temperature ~60°C)
- 3. Wash slides in PBS, 3 x 1 min
- 4. Blocking endogenous peroxidase activity
 - a. Incubate the sections with 3% hydrogen peroxide in PBS (freshly prepared!) for ${\bf 5}\ {\bf min}$
- 5. Wash slides in PBS, 2 x 1 min
- 6. Wash slides in TBST, 1 x 2 min
- 7. **Optional:** Perform Avidin-Biotin-Block according to manufacturer's instructions. *Note:*Certain tissues (e.g. liver, kidney) contain high levels of endogenous biotin. The Avidin-Biotin blocking step is recommended when using the ABC system for these tissues. If the background problem persists, consider trying a polymer-based detection system instead of biotinylated secondary antibody / ABC system.
- 8. Block in Protein Block, Serum-Free for 10 min
- 9. Drain slides (do not rinse)
- 10. Apply primary antibody diluted in Antibody Diluent and incubate in a humidified chamber for 1 h at room temperature

Suggested dilution: 1:1000 in Antibody Diluent

11. Wash slides in TBST, 3 x 2 min

Apply secondary antibody diluted in Antibody Diluent for 30 min at room temperature. *Suggested concentration: 5 μg/ml* *Perform step 13 in the interim*

- 13. Prepare the ABC-reagent: 5 ml PBS + 1 drop A + 1 drop B, incubate for 30 min
- 14. Wash slides in TBST, 3 x 2 min

15. Apply the ABC reagent for 30 min at room temperature

- 16. Wash slides in TBST, 3 x 2 min
- 17. Apply the DAB substrate, 1-10 min

Observe the staining with a microscope! Development times may differ depending upon the level of antigen

- 18. Stop the DAB reaction with deionized water
- 19. Optional: Counterstain
 - a. Follow the manufacturer's instructions for counterstaining and bluing
- 20. Wash slides in deionized water for 1 min

21. Dehydrate tissue sections:

- a. 70% EtOH 2 x 10 sec
- b. 80% EtOH 1 x 10 sec
- c. 90% EtOH 1 x 10 sec
- d. 2-Propanol 2 x 1 min
- e. Xylol 3 x 2 min

22. Mount slides in a suitable organic mounting medium and add coverslip

Note: The SYSY standard protocol generates good results in the SYSY labs and may be used as a reference. However, to achieve the highest

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specific signal and lowest non-specific background signal, the best antigen retrieval condition, antibody concentration, incubation temperature, and incubation time must be determined individually. Please also refer to our general protocols.