

Reference Protocol for biotinylated Anti-Ig kappa light chain (HS-346 011BT) Immunohistochemistry using DAB as Chromogen

Tissue Fixation

- 3.7% formaldehyde (24 h), 3.5 µM paraffin sections

Materials and Reagents

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|--|---------------------|
| • Food Steamer | Braun, Multigourmet |
| • Staining Containers with slide holders (e.g. Tissue-Tek) | |
| • Protein Block, Serum-Free | Agilent X0909 |
| • Antibody diluent | Agilent S2022 |
| • ABC HRP Kit, Standard | Vectorlabs PK-4000 |
| • ImmPACT DAB | Vectorlabs SK-4105 |
| • Hydrogen peroxide 30% | Merck 1.07298.0250 |
| • PBS (pH 7.4) | |
| • TBST (TBS, 0.05% Tween 20, pH 7.6) | |
| • Antigen Retrieval buffer:
Citrate Buffer (10 mM Citrate, 0.05% Tween 20, pH 6.0) | |
| • Xylene, 100% ethanol, 90% ethanol, 80% ethanol and 70% ethanol, 2-propanol | |
| • Optional: Hematoxylin Solution (Mayer's, Modified) or other nuclear counterstain | |
| • Optional: Avidin/Biotin Blocking Kit | Vectorlabs SP-2001 |
| • Non-aqueous mounting medium | |

Method

1. Deparaffinize and hydrate tissue sections

- | | |
|--------------------|------------|
| a. Xylol | 2 x 5 min |
| b. 100% EtOH | 2 x 2 min |
| c. 90% EtOH | 1 x 2 min |
| d. 80% EtOH | 1 x 2 min |
| e. 70% EtOH | 2 x 2 min |
| f. Deionized Water | 1 x 20 sec |
| g. PBS | 1 x 2 min |

*Keep the slides in PBS until ready to perform the Antigen Retrieval.
Do not allow the slides to dry out*

2. Antigen Retrieval (AR) using a food steamer

- a. Heat the steamer with a suitable staining container filled with Antigen Retrieval buffer to **~97°C**
- b. Transfer the sections into the staining box, wait until the temperature reaches **97°C**
- c. Incubate the sections in the steamer for **30 min**
- d. Remove the staining container from the steamer and allow the slides to cool down for **20 min** (target end temperature **~60°C**)

3. Wash slides in PBS, 3 x 1 min

4. Blocking endogenous peroxidase activity

- a. Incubate the sections with 3% hydrogen peroxide in PBS (freshly prepared!) for **5 min**

5. Wash slides in PBS, 2 x 1 min

6. Wash slides in TBST, 1 x 2 min

7. **Optional:** Perform Avidin-Biotin-Block according to manufacturer's instructions.

Note: Certain tissues (e.g. liver, kidney) contain high levels of endogenous biotin. The Avidin-Biotin blocking step is recommended when using the ABC system for these tissues. If the background problem persists, consider trying a polymer-based detection system instead of biotinylated secondary antibody / ABC system.

8. Block in Protein Block, Serum-Free for **10 min**

9. **Drain slides (do not rinse)**

10. **Apply primary antibody diluted in Antibody Diluent** and incubate in a humidified chamber for **1 h at room temperature**

Suggested dilution: 1:1000 in Antibody Diluent

Perform step 11 in the interim

11. **Prepare the ABC-reagent:** 5 ml PBS + 1 drop A + 1 drop B, incubate for 30 min

12. Wash slides in TBST, 3 x 2 min

13. **Apply the ABC reagent for 30 min at room temperature**

14. Wash slides in TBST, 3 x 2 min

15. **Apply the DAB substrate, 1-10 min**

***Observe the staining with a microscope!**

Development times may differ depending upon the level of antigen*

16. Stop the DAB reaction with deionized water

17. **Optional: Counterstain**

- a. Follow the manufacturer's instructions for counterstaining and bluing

18. Wash slides in deionized water for 1 min

19. **Dehydrate tissue sections:**

- a. **70% EtOH** **2 x 10 sec**
- b. **80% EtOH** **1 x 10 sec**
- c. **90% EtOH** **1 x 10 sec**
- d. **2-Propanol** **2 x 1 min**
- e. **Xylol** **3 x 2 min**

20. **Mount slides in a suitable organic mounting medium and add coverslip**

Note: The SYSY standard protocol generates good results in the SYSY labs and may be used as a reference. However, to achieve the highest specific signal and lowest non-specific background signal, the best antigen retrieval condition, antibody concentration, incubation temperature, and incubation time must be determined individually. Please also refer to our general protocols.