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Reference Protocol for Anti-CD39 (HS-493 017) Immunohistochemistry using DAB as Chromogen

Tissue Fixation

+ 3.7% formaldehyde (24 h), 3.5 μM paraffin sections

Materials and Reagents

Food Steamer	Braun, Multigourmet			
Staining Containers with slide holders (e.g. Tissue-Tek)				
Protein Block, Serum-Free	Agilent	X0909		
Antibody diluent	Agilent	S2022		
Biotinylated anti-rat antibody	Jackson	712-065-153		
ABC HRP Kit, Standard	Vectorlabs	PK-4000		
ImmPACT DAB	Vectorlabs	SK-4105		
Hydrogen peroxide 30%	Merck	1.07298.0250		
• PBS (pH 7.4)				
• TBST (TBS, 0.05% Tween 20, pH 7.6)				
 Antigen Retrieval buffer: Citrate Buffer (10 mM Citrate, 0.05% Tween 20, pH 6.0) 				
• Xylene, 100% ethanol, 90% ethanol, 80% ethanol and 70% ethanol, 2-propanol				
Optional: Hematoxylin Solution (Mayer's, Modified) or other nuclear counterstain				
Optional: Avidin/Biotin Blocking Kit	Vectorlabs SP-2001			

Non-aqueous mounting medium

Method

1. Deparaffinize and hydrate tissue sections

a.	Xylol	2 x 5 min
b.	100% EtOH	2 x 2 min
c.	90% EtOH	1 x 2 min
d.	80% EtOH	1 x 2 min
e.	70% EtOH	2 x 2 min
f.	Deionized Water	1 x 20 sec

g. PBS 1 x 2 min

Keep the slides in PBS until ready to perform the Antigen Retrieval. Do not allow the slides to dry out

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2. Antigen Retrieval (AR) using a food steamer

- a. Heat the steamer with a suitable staining container filled with Antigen Retrieval buffer to ~97°C
- b. Transfer the sections into the staining box, wait until the temperature reaches 97°C
- c. Incubate the sections in the steamer for 30 min
- Remove the staining container from the steamer and allow the slides to cool down for 20 min (target end temperature ~60°C)
- 3. Wash slides in PBS, 3 x 1 min
- 4. Blocking endogenous peroxidase activity
 - a. Incubate the sections with 3% hydrogen peroxide in PBS (freshly prepared!) for ${\bf 5}\ {\bf min}$
- 5. Wash slides in PBS, 2 x 1 min
- 6. Wash slides in TBST, 1 x 2 min
- 7. **Optional:** Perform Avidin-Biotin-Block according to manufacturer's instructions. *Note:*Certain tissues (e.g. liver, kidney) contain high levels of endogenous biotin. The Avidin-Biotin blocking step is recommended when using the ABC system for these tissues. If the background problem persists, consider trying a polymer-based detection system instead of biotinylated secondary antibody / ABC system.
- 8. Block in Protein Block, Serum-Free for 10 min
- 9. Drain slides (do not rinse)
- 10. Apply primary antibody diluted in Antibody Diluent and incubate in a humidified chamber for 1 h at room temperature

Suggested dilution: 1:100 in Antibody Diluent

11. Wash slides in TBST, 3 x 2 min

Apply secondary antibody diluted in Antibody Diluent for 30 min at room temperature. *Suggested concentration: 5 μg/ml* *Perform step 13 in the interim*

- 13. Prepare the ABC-reagent: 5 ml PBS + 1 drop A + 1 drop B, incubate for 30 min
- 14. Wash slides in TBST, 3 x 2 min

15. Apply the ABC reagent for 30 min at room temperature

- 16. Wash slides in TBST, 3 x 2 min
- 17. Apply the DAB substrate, 1-10 min

Observe the staining with a microscope! Development times may differ depending upon the level of antigen

- 18. Stop the DAB reaction with deionized water
- 19. Optional: Counterstain
 - a. Follow the manufacturer's instructions for counterstaining and bluing
- 20. Wash slides in deionized water for 1 min

21. Dehydrate tissue sections:

- a. 70% EtOH 2 x 10 sec
- b. 80% EtOH 1 x 10 sec
- c. 90% EtOH 1 x 10 sec
- d. 2-Propanol 2 x 1 min
- e. Xylol 3 x 2 min

22. Mount slides in a suitable organic mounting medium and add coverslip

Note: The SYSY standard protocol generates good results in the SYSY labs and may be used as a reference. However, to achieve the highest

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specific signal and lowest non-specific background signal, the best antigen retrieval condition, antibody concentration, incubation temperature, and incubation time must be determined individually. Please also refer to our general protocols.